

$^{13}\text{C}$  NMR STUDIES OF THE FOUR 20,22-EPOXYCHOLESTEROLS  
AND THE TWO 20(22)-DEHYDROCHOLESTEROLS

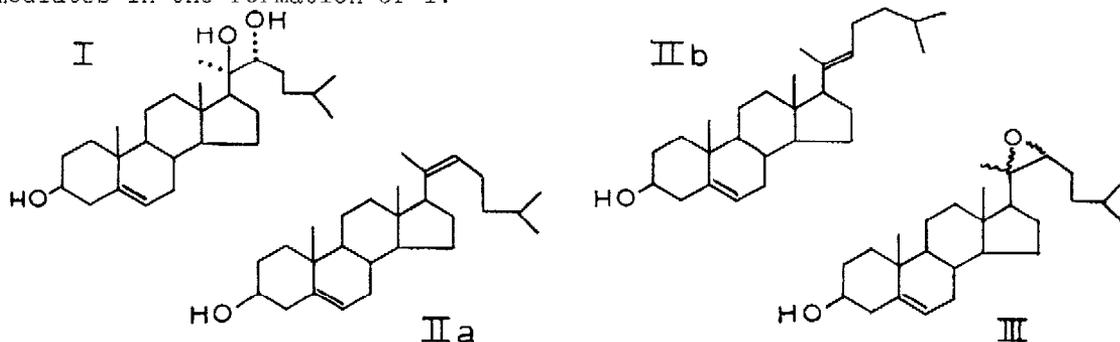
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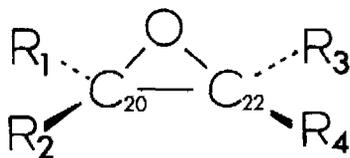
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Recent investigators<sup>1</sup> of the biological conversion of cholesterol to pregnenolone via (20R,22R)-20,22-dihydroxycholesterol (I) have suggested 20(22)-dehydrocholesterol (IIa or IIb) and a 20,22-epoxycholesterol (III) as intermediates in the formation of I.



The stereochemistry of such intermediates is important since a biochemical opening of the epoxide ring would require a specific configuration (which would be formed from only one of the dienes IIa or IIb) in order to give the biologically correct 20R,22R configuration<sup>2</sup> for I.

The  $^{13}\text{C}$  nmr technique has been shown to be useful as a probe of steric interactions of epoxide substituents<sup>3-5</sup>. However, the lack of data on complex tri-substituted acyclic epoxides and our interest in the possible use of  $^{13}\text{C}$  nmr in conformational studies of various cholesterol side chain derivatives have led us to synthesize compounds IIa, IIb and IIIa-d. We wish to report here the synthesis and  $^{13}\text{C}$  nmr data of these compounds.



	<u>R<sub>1</sub></u>	<u>R<sub>2</sub></u>	<u>R<sub>3</sub></u>	<u>R<sub>4</sub></u>
IIIa (20 <u>R</u> ,22 <u>S</u> )	CH <sub>3</sub>	Steroid nucleus	H	C <sub>5</sub> H <sub>11</sub>
IIIb (20 <u>S</u> ,22 <u>R</u> )	Steroid nucleus	CH <sub>3</sub>	C <sub>5</sub> H <sub>11</sub>	H
IIIc (20 <u>R</u> ,22 <u>R</u> )	CH <sub>3</sub>	Steroid nucleus	C <sub>5</sub> H <sub>11</sub>	H
IIId (20 <u>S</u> ,22 <u>S</u> )	Steroid nucleus	CH <sub>3</sub>	H	C <sub>5</sub> H <sub>11</sub>

The <sup>13</sup>C chemical shift assignments of IIa, IIb and IIIa-d have been made and together with those of cholesterol (IV)<sup>10</sup> are listed in the Table. Assignments are based on noise and CW decoupled spectra and on comparison with the cholesterol data.

In view of the planar olefinic structure and the near planarity of the epoxide carbon-substituent bonds, it was expected that some similarities in chemical shift trends within the two series would be observed. From the Table it can be seen in both series that geometrical changes about C-20 - C-22 are reflected primarily in the chemical shifts of C-17, C-18 and C-20 to C-23 with little or no effect apparent in the remaining carbon resonances. The effect of configuration on the shifts of C-17 and C-21 is consistent with previous results<sup>4,9</sup> and the cis shielding influence of the C-23 group is apparent. Thus, C-21 is more highly shielded in those configurations where it is cis to C-23 (IIb, IIIc, IIId) rather than trans (IIa, IIIa, IIIb). A similar influence is apparent in the C-17 data.

While configurational changes about C-20 - C-22 result in a maximum variation of only 1 ppm in the C-20 chemical shift, very substantial differences (6.7 ppm) are seen in the C-22 data. In IIa and IIb the monosubstituted C-22 is shielded relative to the disubstituted C-20, as would be expected from previous work<sup>9</sup>. However, while a similar relationship could be expected for the epoxides<sup>4,5</sup>, the opposite is seen to be the case and in IIIa and IIIb the C-22 (doublet in SFORD) is in fact well downfield of C-20 (singlet in SFORD). In IIIc and IIId C-20 and C-22 have similar shifts. The deshielding effect on C-22 (and possibly the increased shielding of C-18 in IIIc and IIId) may well reflect the influence of the epoxide group on conformational preference about the C-17 - C-20 bond.

TABLE.  $^{13}\text{C}^{\text{a,b}}$  nmr chemical shifts of compounds IIa, IIb, IIIa-d, IV

Carbon	IV	IIa	IIb	IIIa	IIIb	IIIc	IIId
13	42.3	45.9	43.6	42.5	42.4	43.2	42.5
14	56.8	55.9	56.5	56.4	56.2	56.7	56.8
15	24.3	24.7	24.5	24.3	24.7	23.9	24.1
16	27.9	26.3	26.0	26.9	26.3	26.2	26.6
17	56.3	51.2	59.1	53.3	50.5	56.7	56.8
18	11.8	14.0	13.0	14.7	14.4	13.2	13.5
20	35.8	134.2	134.1	61.1	61.0	60.0	60.4
21	18.7	22.8	17.8	22.6	22.7	17.3	20.3
22	36.5	129.6	126.1	66.5	65.1	60.4	59.8
23	23.8	24.7	24.9	23.4	24.7	21.9	22.4
24	39.9	38.5	38.9	36.4	35.8	35.6	35.7

<sup>a</sup>In ppm ( $\pm 0.1$  ppm) downfield from TMS, 0.3M in  $\text{CDCl}_3$ . Spectra were taken in FT mode on a Varian HA100 modified with Digilab NMR-3HC.

<sup>b</sup>Remaining carbon resonances had similar  $\delta$ 's in all seven compounds:

C1 ( $\delta 37.4 \pm 0.1$ ), C2 ( $\delta 32.1 \pm 0.3$ ), C3 ( $\delta 71.8 \pm 0.2$ ), C4 ( $\delta 42.4 \pm 0.1$ ),  
 C5 ( $\delta 141.0 \pm 0.2$ ), C6 ( $\delta 121.6 \pm 0.2$ ), C7 ( $\delta 31.9 \pm 0.2$ ),  
 C8 ( $\delta 31.9 \pm 0.2$ ), C9 ( $\delta 50.4 \pm 0.2$ ), C10 ( $\delta 36.6 \pm 0.2$ ),  
 C11 ( $\delta 21.0 \pm 0.3$ ), C12 ( $\delta 39.6 \pm 0.3$ ), C19 ( $\delta 19.4 \pm 0.1$ ),  
 C25 ( $\delta 28.0 \pm 0.2$ ), C26 ( $\delta 22.6 \pm 0.2$ ), C27 ( $\delta 22.6 \pm 0.2$ )

The synthesis of IIb has already been described<sup>6</sup>. Following the modified method by Josan and Eastwood<sup>7</sup>, the  $3\beta$ -acetate of IIa was synthesized from (20R,22S)- $3\beta,20,22$ -trihydroxycholesterol  $3\beta$ -acetate and the resulting acetate was hydrogenolyzed with LAH to give the corresponding dehydrocholesterol IIa.

Oxidation of the  $3\beta$ -acetate of IIb with *m*-chloroperbenzoic acid gave the  $3\beta$ -acetates of IIIc and IIId which were separated on preparative tlc, followed by hydrogenolysis of their acetates with LAH to give the corresponding epoxy-cholesterols, IIIc and IIId.

(20R)- $3\beta$ -acetoxy-20(2'-tetrahydropyranyloxy)-pregn-5-ene-20-carbonitrile was converted to the corresponding aldehyde by reduction with diisobutylaluminum hydride to the imine, followed by aqueous acetic acid hydrolysis in ethanol. Grignard reaction of this  $3\beta$ -acetate aldehyde with isoamylmagnesium bromide in  $\text{CH}_2\text{Cl}_2$  at  $-70^\circ$  and cleavage of the THP ether under acidic conditions gave the  $3\beta$ -acetate of I which was converted to its 22-mesylate. IIIa was obtained by treatment of this mesylate with aqueous KOH in pyridine.

Oxidation of 3 $\beta$ -acetate of IIb with OsO<sub>4</sub> and hydrolysis of the osmate with aqueous NaHSO<sub>3</sub> yielded the (20S,22S)- and (20R,22R)- glycol acetates which were separated by HPLC. The major fraction (20S,22S) glycol was transformed to IIIb as described above. The full details of these syntheses will be described elsewhere<sup>8</sup>.

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#### References

1. Kraaiipoel, R. J., Degenhart, H. J., Leferink, J. G., van Beek, V., de Leenw-Boon, H., and Visser, H. K. A., FEBS Lett. 50, 204 (1975); Kraaiipoel, R. J., Degenhart, H. J., van Beek, V. and de Leenw-Boon, H., Abeln, G., Visser, H. K. A. and Leferink, J. G., FEBS Lett., 54, (1975); Kraaiipoel, H. J., Degenhart, H. J. and Leferink, J. G., FEBS Lett., 57, 294 (1975).
2. Chaudhuri, N. K., Nickolson, R., Kimball, H. and Gut, M., Steroids, 15, 525 (1970).
3. Easton, N. R., Anct, F. A. L., Burns, P. A. and Foote, C. S., J. Am. Chem. Soc., 96, 3945 (1974).
4. Davis, S. G. and Whitham, G. H. J. Chem. Soc., Perkins II, (1975), 861.
5. Paulson, D. R., Tang, F., Moran, G., Murray, A. S., Pelka, B. and Vasquez, E., J. Org. Chem., 40(2), 184 (1975).
6. Chaudhuri, N. K., Nickolson, R., Williams, J. G., and Gut, M., J. Org. Chem., 34, 3767 (1969).
7. Josan, J. S. and Eastwood, F. W., Aust. J. Chem., 21, 2013 (1968).
8. Byon, C. Y. and Gut, M. in preparation.
9. Stothers, J. B. "Carbon-13 NMR Spectroscopy", Academic Press, New York, N.Y. (1972).
10. Letourneux, Y., Khuong-Huu, Q., Gut, M. and Lukacs, G., J. Org. Chem. 40, 1674 (1975).